

MOLab TqPCR Protocol (Using 2x SYBR Green Master Mix) (TCH @ 08/07/2017)

Preparation of qPCR primers

- Design with Primer3 Plus: <http://primer3plus.com/cgi-bin/dev/primer3plus.cgi>
- qPCR primer features: 18mers; T_m=60°C; amplicon size: 100-200bps
- **Dilute qPCR primers:** dilute Forward (Fwd) and Reverse (Rev) primers with ddH₂O to make 1 µg/µl stock solution; take 5 µl from each primer stock solution (total = 10 µl) and add 490 µl ddH₂O; mix well and keep at -20°C or -80°C for qPCR use (*i.e.*, each primer @ 10ng/µl).
- 2x SYBR Green qPCR mix: Bio-Rad, Thermo Fisher brands; We are using Bimake's 2x SYBR Green qPCR Master Mix (Cat# B21204)

qPCR Reaction Setup and TqPCR Cycling Program

RT-PCR (cDNA) template:	2.5 µl
Primer mix (each at 10ng/µl):	2.5 µl
2x SYBR Green Master Mix:	5.0 µl (or 2.5 µl to save \$\$\$)**

- Add 10 µl mineral oil/tube or well
- Seal plates with film
- Briefly spin/tap down the contents in each tube/well
- Run **Touchdown-qPCR** program as follows (on Bio-Rad CFX-Connect unit):

1. 95°C × 3' for one cycle
2. 95°C × 20"
3. 66°C × 10"
4. GOTO **Step #2** for 4 cycles by decreasing 3°C per cycle
5. 95°C × 20"
6. 55°C × 10"
7. 70°C × 1"
8. plate read
9. GOTO **Step #5** for 40 cycles

**** We found that 2.5 µl of 2x Master Mix yielded similar C_q values (to that of 5.0 µl 2x Master mix) for transcripts with average or above average abundance. Nonetheless, the use of 5.0 µl of 2x SYBR Green Master Mix is recommended to detect extremely low abundant transcripts.**

Reference

Zhang Q, Wang J, Deng F, Yan Z, Xia Y, Wang Z, et al. (2015) TqPCR: A Touchdown qPCR Assay with Significantly Improved Detection Sensitivity and Amplification Efficiency of SYBR Green qPCR. PLoS ONE 10(7): e0132666. doi:10.1371/journal.pone.0132666
<http://journals.plos.org/plosone/article?id=10.1371/journal.pone.0132666>