

# General Protocol for Running the Universal TaqMan qPCR Analysis

(Hao Wang @ 08/01/2021; edited by TCH)

## PART 1: Reverse Transcription

**Step 1:** Prepare the first step reaction (**RT Mix1**) as described in the following table:

	Volume
Total RNA	5~10 $\mu$ l (usually 1~5 $\mu$ g total RNA)
<b>TMP-RT primer mix</b>	2 $\mu$ l
H <sub>2</sub> O	Varied vol. to make total vol. of 12 $\mu$ l
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Total	12 $\mu$ l

Run the first step reaction (**RT Mix1**) (**program RT1**) as the following: 70°C, 5min, place **RT Mix1** on ice

**Step 2:** Prepare **RT Mix2** as described in the following table:

	Volume
10x RT Buffer	2.5 $\mu$ l
dNTPs (5mM)	1 $\mu$ l
H <sub>2</sub> O	4.5 $\mu$ l
RT enzyme (M-MuLV)	0.2 $\mu$ l
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Total	8.2 $\mu$ l

**Step 3:** Mix reactants in Steps #1 and #2 as described in the following table:

	Volume
RT mix1-Step 1	12 $\mu$ l
RT mix2-Step 2	8.2 $\mu$ l
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Total	~20 $\mu$ l

Run the reaction (**program RT**): 37°C, 60min; Then heat the reaction mix at 92°C, 1min; 12°C,  $\infty$ .

Add 80 $\mu$ l ddH<sub>2</sub>O after RT reaction, store the sample at -80°C (=cDNA stock solution).

Template for qPCR: 10  $\mu$ l cDNA/RT product + 490  $\mu$ l ddH<sub>2</sub>O (=1:500 dilution; higher dilutions may be needed)

**NOTE: It is important to keep RT products at -80C; Aliquot if possible to avoid repeated thaws.**

## PART 2: Universal TaqMan qPCR Reaction

Set up each reaction tube as described in the following on a cooler (in triplicated in general):

	Volume
<b>2x UQE2 Master Mix</b>	5 $\mu$ l
Forward Primer (10ng/ $\mu$ l)	2 $\mu$ l
Template	2 $\mu$ l
<b>TPRP Mix</b>	1 $\mu$ l
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Total	10 $\mu$ l

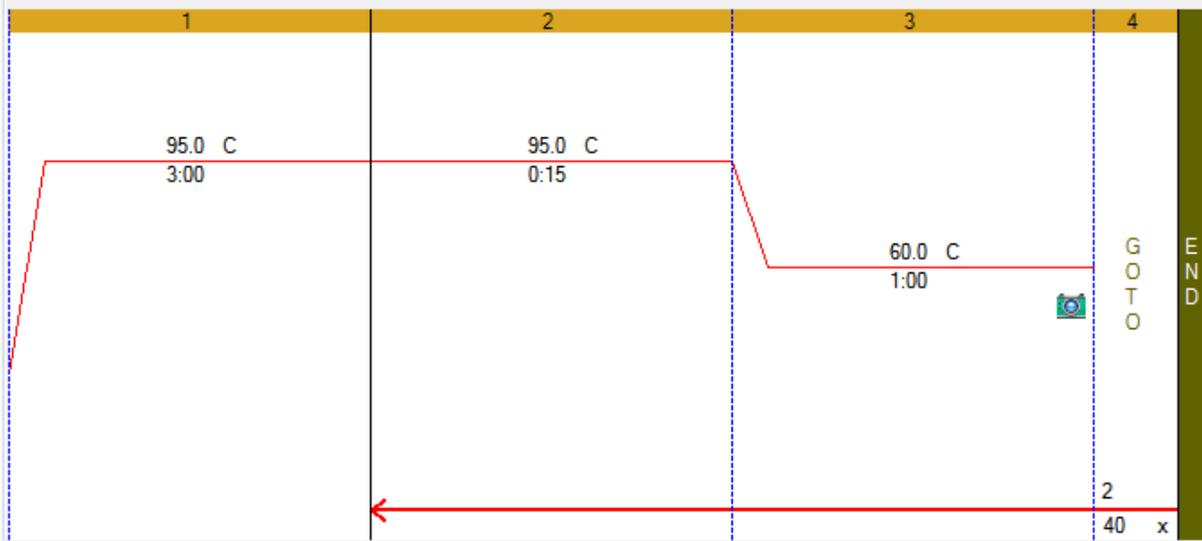
Launch the qPCR program (**TaqMan-2021** on the desktop) as described in the following:

Step 1: 95°C, 3min

Step 2: 95°C, 15sec

Step 3: 60°C, 1min, go to Step 2, 39x

Protocol: Taqman-2021.prcl



1	95.0 C for 3:00
2	95.0 C for 0:15
3	60.0 C for 1:00
	+ Plate Read
4	GOTO 2 , 40 more times
	END