DNA Sample and primer Preparation for DNA sequencing

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- 1. DNA sample preparation:
 - Take 50µl of your Wizard miniprep DNA (*see Protocol for Wizard Plus minipreps*) and re-precipitate by ethanol and 7.5 M NH4OAc (*see Protocol for ethanol precipitation of DNA sample*).
 - Wash twice with 70% ethanol and dissolve the pallet into 10µl ddH20 (<u>make</u> sure you wash the pallets very well because even 10-20 mM salt can inhibit the sequencing reaction! The washing process can also remove some inhibitors). You need to check 1ul on an agarose gel.
- 2. Sequencing Primer preparation:
 - Provide at least 10µl sequencing primer (you will need 1-2ul per sequencing reaction) at the concentration of 30 ng/µl.
 - Too high concentration of primer might increase the background from non-specific priming.
- Label each tube well and send for sequencing (along with a DNA Sequencing <u>Request From; ask T.-C. for this form</u>). The sample drop place for University of Chicago Cancer Research Center DNA Sequencing facility is located at <u>Cummings 10th floor hallway</u>. Put into the freezer and sign up the check-in sheet.
- 4. The turnaround time is usually 24hrs. Follow Protocol A38 to download and analyze the sequencing data.

Reference: http://cancer-seqbase.uchicago.edu